

Sialic acids: biomarkers in endocrinal cancers

Shyamasree Ghosh

Received: 15 January 2015 / Revised: 11 February 2015 / Accepted: 18 February 2015 / Published online: 17 March 2015
© Springer Science+Business Media New York 2015

Abstract Sialylations are post translational modification of proteins and lipids that play important role in recognition, signaling, immunological response and cell-cell interaction. Improper sialylations due to altered sialyl transferases, sialidases, gene structure and expression, sialic acid metabolism however lead to diseases and thus sialic acids form an important biomarker in disease. In the endocrinal biology such improper sialylations including altered expression of sialylated moieties have been shown to be associated with disorders. Cancer still remains to be the major cause of global death and the cancer of the endocrine organs suffer from the dearth of appropriate markers for disease prediction at the early stage and monitoring. This review is aimed at evaluating the role of sialic acids as markers in endocrinal disorders with special reference to cancer of the endocrine organs. The current study is summarized under the following headings of altered sialylations in endocrinal cancer of the (i) ovary (ii) pancreas (iii) thyroid (iv) adrenal and (v) pituitary gland. Studies in expression of sialic acid in testis cancer are limited. The future scope of this review remains in the targeting of endocrinal cancer by targeting altered sialylation which is a common expression associated with endocrinal cancer.

Keywords Sialic acids · Marker · Endocrines · Cancer · Sialylations · Sialoglycoconjugate

Introduction

Sialic acids are negatively charged nine-carbon carboxylated monosaccharides on glycosylated proteins and lipids formed due to post translational modification. Located terminally on the cell surface they constitute the glycocalyx, forming receptors that contribute to the structure, function, recognition, signaling, and interaction of the host cell with other molecules and is vital for cell-cell and cell-pathogen interaction. A battery of enzymes including (i) sialyltransferases catalyzing their biosynthesis, (ii) sialidases also termed as neuraminidases, catalyzing their cleavage regulate their occurrence in the mammalian system [1]. Studies in infection by *Trypanosoma cruzi*, reveal that signaling by sialylated mucins *via* sialic acid-binding Ig-like lectin receptors enable inhibitory effects on CD4+ T cells causing G1 cell cycle arrest thereby modulating host immune responses [2]. Sialylation of Fc region of antibodies have been reported to show anti-inflammatory properties on one hand and on the other hand over-expression of sialylation affects antibody function by altering the antigen specific binding, decreased Fc mediated effector functions and increased protease sensitivity [3]. Polysialic acids (PSA) linked by (α 2-8) glycosidic linkages by the enzyme polysialyltransferase-1 (ST8SiaIV) and sialyltransferase-X (STX or ST8SiaII) is a normal constituent of the human neuronal cell adhesion molecule (NCAM) that play role in neural development in processes including neuronal regeneration, neurite outgrowth, axonal growth, and neuronal sprouting [4]. Sialylations contribute to normal function of ion channels [5] and have been shown to vary in an “M” shaped manner during normal menstrual cycle of healthy women [6]. Upregulation of sialylation of apolipoprotein C-III (apoCIII) protein coded by APOC3 gene, which is a risk factor in cardiovascular diseases has been demonstrated for

S. Ghosh (✉)

School of Biological Sciences, National Institute of Science Education and Research (NISER), under Dept of Atomic Energy (DAE), Government of India, Institute of Physics Campus, Sachivalaya Marg, PO. Sainik School, Bhubaneswar 751 005, India
e-mail: sree.s@niser.ac.in
e-mail: shyamasree_b@yahoo.com

its proinflammatory properties in type 2 diabetic patients [7] while enhanced sialylation have been reported during hypo and hyperthyroidism [8]. Sialylation of thyrotropin (TSH) hormone has been reported to increase the hormone bioactivity [9].

Altered sialylations and their modifications including acetylation of sialylated proteins [10–15], together with the altered sialyl transferases [16, 17] are associated with disorders and have been reported as disease markers and potential targets for therapeutic approaches. Lysosomal neuraminidase deficiency, including sialidosis and galactosialidosis leads to genetical storage disorders [18]. In cancer, altered sialic acid profile plays dominant role enhancing tumor growth, metastasis, evading immune surveillance, escaping the apoptotic pathways, leading to cancer cell survival and resistance to therapy [10–15]. Overexpression of *O*-acetylated sialoglycoconjugates in childhood acute lymphoblastic leukemia (ALL) has been documented in correlating the monitoring of cancer progression and enabling cancer cell survival [10, 11, 14, 15] establishing their potential biological role in cancer progression.

Overexpression of α 2,6 sialylation of glycoproteins due to expression of increased ST6GalII (β -galactoside- α 2,6 sialyltransferase) have been reported to contribute to metastasis, increase migration by increased sialylation of integrin β 1 and overexpression of CDw75 with Sia- α (2-6) Gal β (1-4) GlcNAc sequence, leading to resistance in colorectal cancer [19, 20]. ST3GalI has been reported of promoting tumor in breast cancer [21].

Increased sialylations together with altered enzyme profile regulating sialylation has been reported in several cancers including childhood acute lymphoblastic leukemia [10–15], colorectal [19, 20], breast [21], colon [22–24], cervical [25], ovary [26–40], prostate cancer [41, 42], pancreatic cancer [43–54], multiple myeloma [55–57], hepatocellular carcinoma [58].

Deletion in the human *cmah* gene coding for the enzyme cytidine monophospho-*N*-acetyl-neuraminic acid hydroxylase (CMP-Neu5Ac hydroxylase) prevents them from synthesizing *N*-glycolyl-neuraminic acid (Neu5Gc). However, in malignant states, Neu5Gc-sialoconjugates, are reported in particular from the gangliosides. Therefore the impact of elevated sialylation in cancers can thus further be appreciated by the observation of occurrence of Neu5Gc in cancer cell lines although they are not reported from adult healthy individuals [59] inferring at the altered sialylation synthetic machinery associated with malignancy.

In the current review the scope of sialylation as disease marker for endocrinal disorders are evaluated and studied under the headings of sialylations in the disorders of the endocrine organs including (i) ovary (ii) pancreas (iii) thyroid (iv) adrenal gland (v) pituitary and (vi) hypothalamus. The future scope of this review remains in the targeting of

endocrinal disorders by targeting altered sialylations, which is a common manifestation in endocrinal disorders.

Sialylations in endocrinal disorders with special reference to cancers

Ovary

Ovarian cancer being lethal when diagnosed late suffers from the limitation of suitable sensitive biomarkers for early tumour detection, inflammation and progression. Increased sialylation ([26–40], Fig. 1) and branching of sialylation mediated by over-expression of sialyl transferase branching enzyme MGAT5 [27], alterations in DNA methylation [28], differential overexpression of sialoglycoproteins in serum [29] and overexpression of ST6GalII (β -galactosamide α 2,6-sialyltransferaseI) gene, and overall increased sialylations in the proximal fluid from patients suffering from ovarian cancer [30], altered mRNA expressions of α 2,3-sialyltransferase ST3GalI, ST3GalIII, ST3GalIV, ST3GalVI, and α 2,6-sialyltransferase ST6GalII together with increased expression of ST3GalI leading to increased α (2-3)-linked sialylation [31] causing increase in tumour sizes, metastasis and resistance to therapy thereby indicating the role of sialic acid as biomarker for ovarian cancer detection and monitoring [26, 29, 30]. Modification of kallikrein 6 (KLK6) with α (2-6)-linked sialic acid in ovarian cancer patients further correlates with the fact that sialic acid could be a potential marker in ovarian cancer [32]. Over expression of sialylation and its branching correlates with inflammation and progress of ovarian tumour [27]. Ascites, plasma and serum from ovarian cancer patients reveal increased sialic acids [33], disialic acids [34], gangliosides [35], altered sialylation of alpha-1 protease inhibitor [36], increased lipid associated sialic acid (LSA, [38–40]) and increased sialylation in the cyst and peritoneal fluids of patients with ovarian cysts and tumors [40] are reported features in ovarian tumours.

Pancreas

Studies with pancreatic adenocarcinoma cell lines reveal that overexpression of α (2-6)-sialic acids that mediate increased adhesion to extracellular matrix (ECM) while overexpressed α (2-3)-sialic acids contribute to increased migration of cells thus overall leading to metastatic effects in cancer [43]. The contributory factors to increased sialylation and sialic acid in pancreatic carcinoma have been attributed to major factors including altered enzymes for biosynthesis of sialic acids [44], overexpression of GD3 synthase thereby inhibiting survival and angiogenesis of pancreatic cancer cells through cell cycle arrest [44], increased serum sialic acids [45] increased cell surface expression of sialic acid modulating migration and matrix adhesion in pancreatic adenocarcinoma [46], increased

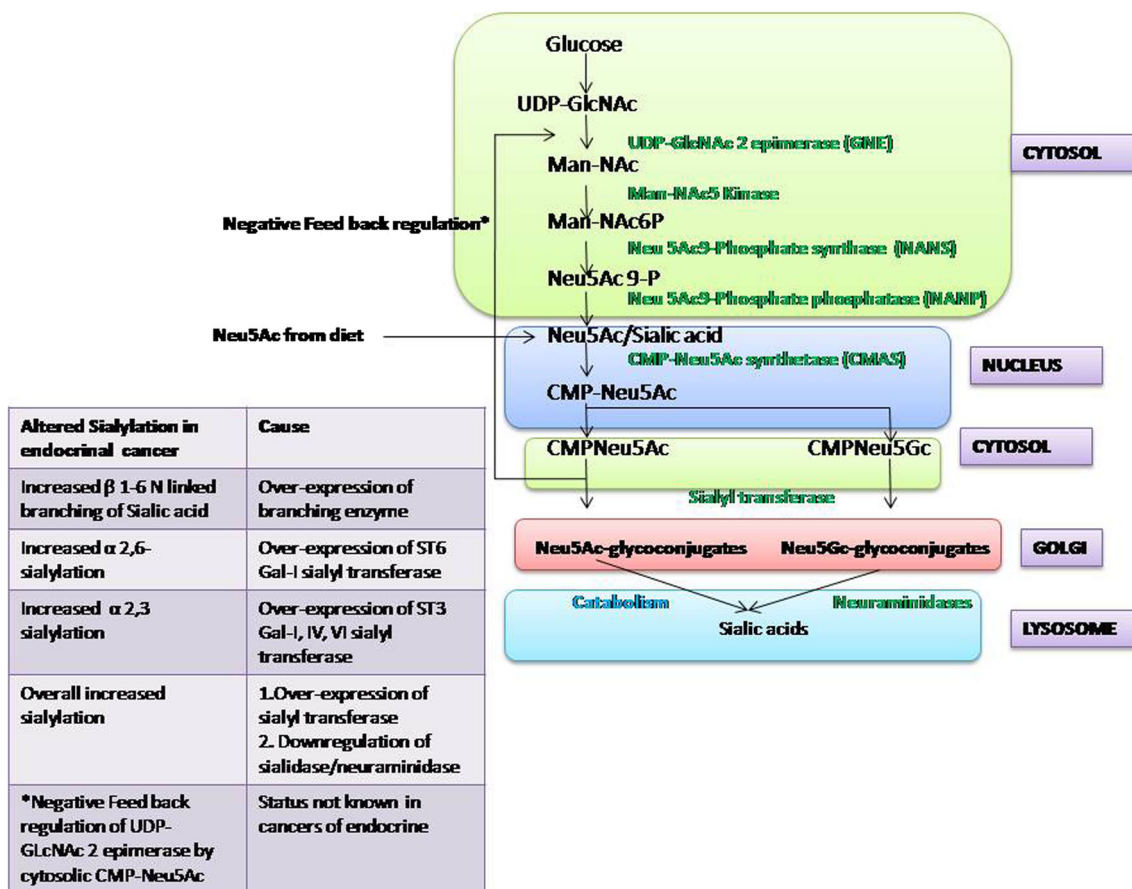


Fig. 1 Sialic acid metabolism and cancer of the endocrine organs. Sialylated glycoconjugates are synthesized from sialic acid/Neu5Ac. It starts with (i) conversion of glucose to UDP-*N*-acetylglucosamine (UDP-GlcNAc) (ii) conversion of UDP-GlcNAc to *N*-acetylmannosamine (ManNAc) by the enzyme UDP-GlcNAc 2 epimerase (GNE) (iii) phosphorylation of ManNAc to ManNAc-6-phosphate by the enzyme ManNAc5 kinase. (iv) ManNAc-6-P is then converted into NeuAc-9-phosphate by the enzyme sialic acid 9-phosphate synthase (NANS) (v) NeuAc-9-P converts into Neu5Ac mediated by sialic acid 9-phosphate phosphatase (NANP). The formed *N*-acetyl neuraminic acid (Neu5Ac)/sialic acid formed in the cytosol then moves into the nucleus through a nuclear pore and is converted into CMP-Neu5Ac by the enzyme CMP-Neu5Ac synthase (CMAS). The enzymatic activity of UDP-GlcNAc2 epimerase is negatively regulated by feedback inhibition by cytosolic CMP-Neu5Ac in normal individuals. However the status of this

regulation is not known in endocrinal cancers. CMP-Neu5Ac enters the cytosol from the nucleus by means of nuclear pores entering the Golgi apparatus where they are catalysed by sialyltransferases (ST) enabling their linkage to glycoconjugated moieties leading to the formation of sialoglycoconjugates. *N*-glycolyl neuraminic acid (Neu5Gc) absent in adult human is, however, observed in cancer mostly entering the body by dietary intake [59]. Neuraminidases/Sialidases catalytically removes the sialic acid from the sialoglycoconjugates. In the endocrine cancer biology a common pattern of overexpression of sialyltransferases, and downregulation of sialidases are reported leading to overall increase in sialylations [1–77] while the status of negative feed back regulation of GNE producing UDP-GlcNAc2 epimerase by CMP-Neu5Ac, in the sialic acid biosynthesis pathway reported from all eukaryotes remains unknown in cancers of the endocrine

expression of α 2,3 sialyltransferase ST3GalIV promoting migration and metastasis in pancreatic adenocarcinoma [46], overall increased sialic acid conjugated *N*-glycans [47], sialylation of MUC1 mucin are reported from benign pancreas and pancreatic duct adenocarcinoma [48]. Alpha2,8-polysialyltransferase enzymes namely ST8Sia IV, ST8Sia II, and ST8Sia III are reported to be capable of incorporating both oligosialic and polysialic acid on various sialylated *N*-acetylglucosaminyl oligosaccharides, including NCAM of which ST8Sia IV and ST8Sia II were reported to efficiently catalyze polysialylation of NCAM as compared to ST8Sia III [4, 49]. Highly polysialylated neural cell adhesion molecule (NCAM) termed as PSA-NCAM have been reported to be a

key molecule initiating metastatic pathway leading to peri neural invasion (pni) [49, 50], observed specially in pancreatic cancer mediated by the cancer cell invasion of nerves. However the complete mechanism of pni remains yet to be deciphered [50]. Down-regulation of two enzymes in the biosynthetic pathway of sialic acid biosynthesis including UDP-*N*-acetylglucosamine 2-epimerase/*N*-acetylmannosamine kinase (GNE) and *N*-acetylneuraminic acid 9-phosphate synthase by tumour suppressor, p16(INK4a) in pancreatic carcinoma model indicates further the correlation of sialylation with cancerous state in pancreatic cancer [51], increased metabolic influx of sialic acids [52], alteration in gene expression [53], overexpression of alpha ST3Gal III [54] are phenotypic

expression in pancreatic cancer with biological function in pancreatic cancer cell progression and metastasis. Bulk metabolic flux through the sialic acid pathway causing about 40 % of proteins with doubled sialylation has been shown to correlate with the altered adhesive properties of pancreatic cancer cells implicating their role in metastasis [52], thus highlighting the role of sialic acid as diagnostic marker in pancreatic cancer.

Diabetes mellitus patient are reported to reveal increased plasma sialic acid content [60] correlating with marked insulin deficiency, increased sialylation of apolipoprotein E [61], glycosylation and sialylation of insulin-like growth factor-binding protein (IGFBP-3). This increases the affinity of IGFBP-3 for IGF-I while sialylation decreases it leading to complications [62]. Sialidase activity of peripheral mononuclear leukocytes in diabetic individuals has been reported to be significantly decreased as compared to normal individuals [63] and decreased glycophorin sialylation leads to increased erythrocyte aggregation that in turn leads to pathogenesis of vascular disease in diabetes [64]. Individuals with gestational diabetes mellitus (GDM), revealed higher placental sialidase activity accounting for reduced sialic acid content of glycodelin-A(GdA) leading to defective immunomodulatory effects in GDM pregnancies [65].

Thyroid

Increased mRNA level expression and activity of thyroid sialyltransferase-I are reported in Graves' disease [66] with increased expression of $\alpha(2-3)$ -sialic acids as compared to $\alpha(2-6)$ -sialic acids in autoimmune thyroiditis [67]. Sialylation has been reported to modulate the bioactivity of thyroid hormones. The impaired thyroid hormone action, and secretion of high amounts of sialylated TSH isoforms in hypothyroid or fetuses showing resistance to thyroid hormone (RTH), has been attributed to early expression of α -2,6-sialyltransferase activity thereby increasing terminal sialylation within thyrotropes [68].

Contrasting reports of sialylation profile exists for malignancy of the thyroid [69, 70]. While some studies have revealed correlation of increased level of $\alpha(2-3)$ -sialic acids with malignant transformation in the thyroid gland [71], decreased sialylation has also been reported to be correlate with the thyroid malignancy [72]. The increased expression of sialylated fibronectin [71] sialic acid masked Lewis(a) antigen [72], serum and overall total sialic acid [73] are reported in human thyroid cancer.

Adrenal cancer

Very few reports exist over the expression of total sialic acid in adrenal cancer. Adrenal cancer reveals overall increase in sialylation as revealed from its total cellular and cytoplasmic

sialic acid content [74] and in plasma bound form [75]. No reports on the sialylation content individually on the glycoproteins and glycolipids are known in studies from adrenal cancer.

Pituitary

Few studies on pituitary and brain cancer reveal altered sialylation, polysialylation of neural cell adhesion molecule [76, 77], serum sialic acids [78, 79] and finds importance as prognostic markers in pituitary cancer [78].

Discussions

Endocrinal disorders and cancers are life threatening. Sialic acids have been reported to be markers in cancer [10–81]. Sialic acid with modification like *O*-acetylations has been reported to play a dominant role in the biology of cancer metastasis and progression. [10–15, 80, 81]. *O*-acetylation of sialic acids have been reported to enable survival of lymphoblasts in childhood acute lymphoblastic leukemia (ALL, 10). 9-*O*-acetylated sialic acids are reported as immunological tool for monitoring pediatric patients suffering from acute lymphoblastic leukemia (ALL, 15). A decrease in the level of *O*-acetylated sialic acids leading to an increased level of expression of sialyl Lewis(X), a tumor-associated antigen have been reported to be associated with cancer metastasis in colorectal carcinoma, [80, 81].

An overall alteration in the sialic acid synthesis machinery has been reported from different endocrinal cancers [26–40, 44–54, 69–79]. However, the status of negative feed back regulation of GNE producing UDP-GlcNAc2 epimerase by CMP-Neu5Ac, in the sialic acid biosynthesis pathway (Fig. 1) reported from all eukaryotes [59] remains yet to be investigated in cancers of the endocrine.

Based on ongoing researches across the globe, the current review highlights that increased sialylation due to altered genetic and metabolic events is a (i) hall mark in the biology of endocrinal cancer and finds application as a marker for detection, monitoring and progress of endocrinal cancer, which suffers from the current limitation of sensitive markers for early detection of the disease and (ii) is a strong indicator of endocrinal disorders. Although reports of sialylation in endocrinal cancers are reported, no reports of additional modifications like *O*-acetylations exists from cancers of the endocrines. This review is the first of its kind to study the increased sialylation in endocrinal disorder with special reference to cancers of the endocrine. This finding has a major impact on the diagnosis, therapeutics and targeting of the endocrinal cancer. The significance of the findings remains that although non specific, but an overall rise in the sialic acid level and sialylated moieties and their regulating enzymes and genes,

in the body, could be early indications of endocrinal cancer, thus strongly suggesting the role of sialic acid in endocrinal cancer prediction, diagnosis and prognosis. The future scope of this review remains that since sialic acid is a common marker observed to be expressed in different types of endocrinal cancer, is it possible to designate sialic acids as a marker to endocrinal cancer and employ it in clinics for early detection of the disease and secondly to target the sialic acid in such cancers thereby proving promising as therapeutic targets in endocrine cancers.

Acknowledgements The author acknowledges School of Biological Sciences, National Institute of Science Education and Research (NISER), Bhubaneswar, under Dept of Atomic Energy (DAE), Govt of India..

Financial support No financial support or grant was taken to complete this work.

Conflict of interest The author discloses no potential conflicts of interest.

References

- Pshezhetsky, A.V., Ashmarina, L.I.: Desialylation of surface receptors as a new dimension in cell signaling. *Biochemistry (Mosc)* **78**, 736–745 (2013)
- Morrot A.: The role of sialic acid-binding receptors (Siglecs) in the immunomodulatory effects of trypanosoma cruzi. *Sialoglycoproteins on the protective immunity of the host. Scientifica (Cairo)* 2013, Article ID 965856, 1–7 (2013)
- Raju, T.S., Lang, S.E.: Diversity in structure and functions of antibody sialylation in the Fc. *Curr. Opin. Biotechnol.* **30**, 147–152 (2014)
- Angata, K., Suzuki, M., McAuliffe, J., Ding, Y., Hindsgaul, O., Fukuda, M.: Differential biosynthesis of polysialic acid on neural cell adhesion molecule (NCAM) and oligosaccharide acceptors by three distinct alpha 2,8-sialyltransferases, ST8Sia IV (PST), ST8SiaII (STX), and ST8SiaIII. *J. Biol. Chem.* **275**, 18594–18601 (2000)
- Baycin-Hizal, D., Gottschalk, A., Jacobson, E., Mai, S., Wolozny, D., Zhang, H., Krag, S.S., Betenbaugh, M.J.: Physiologic and pathophysiological consequences of altered sialylation and glycosylation on ion channel function. *Biochem. Biophys. Res. Commun.* **453**, 243–253 (2014)
- Wide, L., Eriksson, K.: Dynamic changes in glycosylation and glycan composition of serum FSH and LH during natural ovarian stimulation. *Ups. J. Med. Sci.* **118**, 153–164 (2013)
- Hiukka, A., Ståhlman, M., Pettersson, C., Levin, M., Adiels, M., Teneberg, S., Leinonen, E.S., Hultén, L.M., Wiklund, O., Oresic, M., Olofsson, S.O., Taskinen, M.R., Ekroos, K., Borén, J.: ApoCIII-enriched LDL in type 2 diabetes displays altered lipid composition, increased susceptibility for sphingomyelinase, and increased binding to biglycan. *Diabetes* **58**, 2018–2026 (2009)
- Nowosadzka, E., Szymonik-Lesiuk, S., Kurzepa, J.: The effects of hypo- and hyperthyroidism on nuclear, cytosolic, endoplasmic and mitochondrial fractions of sialoglycoproteins in rabbit hepatocytes. *Folia Biol. (Praha)* **55**, 7–10 (2009)
- Trojan, J., Theodoropoulou, M., Usadel, K.H., Stalla, G.K., Schaaf, L.: Modulation of human thyrotropin oligosaccharide structures—enhanced proportion of sialylated and terminally galactosylated serum thyrotropin isoforms in subclinical and overt primary hypothyroidism. *J. Endocrinol.* **158**, 359–365 (1998)
- Ghosh, S., Bandyopadhyay, S., Mukherjee, K., Mallick, A., Pal, S., Mandal, C., Bhattacharya, D.K., Mandal, C.: O-acetylation of sialic acids is required for the survival of lymphoblasts in childhood acute lymphoblastic leukemia (ALL). *Glycoconj. J.* **24**, 17–24 (2007)
- Ghosh, S., Bandyopadhyay, S., Mallick, A., Pal, S., Vlasak, R., Bhattacharya, D.K., Mandal, C.: Interferon gamma promotes survival of lymphoblasts overexpressing 9-O-acetylated sialoglycoconjugates in childhood acute lymphoblastic leukaemia (ALL). *J. Cell. Biochem.* **95**, 206–216 (2005)
- Ghosh, S., Bandyopadhyay, S., Pal, S., Das, B., Bhattacharya, D.K., Mandal, C.: Increased interferon gamma production by peripheral blood mononuclear cells in response to stimulation of overexpressed disease-specific 9-O-acetylated sialoglycoconjugates in children suffering from acute lymphoblastic leukaemia. *Br. J. Haematol.* **128**, 35–41 (2005)
- Ghosh, S., Bandyopadhyay, S., Bhattacharya, D.K., Mandal, C.: Altered erythrocyte membrane characteristics during anemia in childhood acute lymphoblastic leukemia. *Ann. Hematol.* **84**, 76–84 (2005)
- Pal, S., Ghosh, S., Bandyopadhyay, S., Mandal, C., Bandyopadhyay, S., Bhattacharya, D.K., Mandal, C.: Differential expression of 9-O-acetylated sialoglycoconjugates on leukemic blasts: a potential tool for long-term monitoring of children with acute lymphoblastic leukemia. *Int. J. Cancer* **111**, 270–277 (2004)
- Pal, S., Ghosh, S., Mandal, C., Kohla, G., Brossmer, R., Isecke, R., Merling, A., Schauer, R., Schwartz-Albiez, R., Bhattacharya, D.K., Mandal, C.: Purification and characterization of 9-O-acetylated sialoglycoproteins from leukemic cells and their potential as immunological tool for monitoring childhood acute lymphoblastic leukemia. *Glycobiology* **14**, 859–870 (2004)
- Dall’Olio, F., Malagolini, N., Trinchera, M., Chiricolo, M.: Sialosignaling: sialyltransferases as engines of self-fueling loops in cancer progression. *Biochim. Biophys. Acta* **1840**, 2752–2764 (2014)
- Büll, C., Stoel, M.A., den Brok, M.H., Adema, G.J.: Sialic acids sweeten a tumor’s life. *Cancer Res.* **74**, 3199–3204 (2014)
- Rottier, R.J., Bonten, E., d’Azzo, A.: A point mutation in the neu-1 locus causes the neuraminidase defect in the SM/J mouse. *Hum. Mol. Genet.* **7**, 313–321 (1988)
- Park, J.J., Lee, M.: Increasing the α 2, 6 sialylation of glycoproteins may contribute to metastatic spread and therapeutic resistance in colorectal cancer. *Gut Liver* **7**, 629–641 (2013)
- Costa-Nogueira, C., Villar-Portela, S., Cuevas, E., Gil-Martín, E., Fernández-Briera, A.: Synthesis and expression of CDw75 antigen in human colorectal cancer. *BMC Cancer* **9**, 1–10 (2009)
- Picco, G., Julien, S., Brockhausen, I., Beatson, R., Antonopoulos, A., Haslam, S., Mandel, U., Dell, A., Pinder, S., Taylor-Papadimitriou, J., Burchell, J.: Over-expression of ST3Gal-I promotes mammary tumorigenesis. *Glycobiology* **20**, 1241–1250 (2010)
- Lee, M., Park, J.J., Ko, Y.G., Lee, Y.S.: Cleavage of ST6Gal I by radiation-induced BACE1 inhibits golgi-anchored ST6Gal I-mediated sialylation of integrin β 1 and migration in colon cancer cells. **7**, 1–10, (2012)
- Park, J.J., Yi, J.Y., Jin, Y.B., Lee, Y.J., Lee, J.S., Lee, Y.S., Ko, Y.G., Lee, M.: Sialylation of epidermal growth factor receptor regulates receptor activity and chemosensitivity to gefitinib in colon cancer cells. *Biochem. Pharmacol.* **83**, 849–857 (2012)
- Swindall, A.F., Bellis, S.L.: Sialylation of the Fas death receptor by ST6Gal-I provides protection against Fas-mediated apoptosis in colon carcinoma cells. *J. Biol. Chem.* **286**, 22982–22990 (2011)
- Kim, H.J., Kim, S.C., Ju, W., Kim, Y.H., Yin, S.Y., Kim, H.J.: Aberrant sialylation and fucosylation of intracellular proteins in cervical tissue are critical markers of cervical carcinogenesis. *Oncol. Rep.* **31**, 1417–1422 (2013)
- Saldova, R., Wormald, M.R., Dwek, R.A., Rudd, P.M.: Glycosylation changes on serum glycoproteins in ovarian cancer

- may contribute to disease pathogenesis. *Dis. Markers* **25**, 219–232 (2008)
27. Saldo, R., Piccard, H., Pérez-Garay, M., Harvey, D.J., Struwe, W.B., Galligan, M.C., Berghmans, N., Madden, S.F., Peracaula, R., Opdenakker, G., Rudd, P.M.: Increase in sialylation and branching in the mouse serum N-glycome correlates with inflammation and ovarian tumour progression. *PLoS ONE* **8**, e71159 (2013)
 28. Saldo, R., Dempsey, E., Pérez-Garay, M., Mariño, K., Watson, J.A., Blanco-Fernández, A., Struwe, W.B., Harvey, D.J., Madden, S.F., Peracaula, R., McCann, A., Rudd, P.M.: 5-AZA-2'-deoxycytidine induced demethylation influences N-glycosylation of secreted glycoproteins in ovarian cancer. *Epigenetics* **6**, 1362–1372 (2011)
 29. Wu, J., Xie, X., Nie, S., Buckanovich, R.J., Lubman, D.M.: Altered expression of sialylated glycoproteins in ovarian cancer sera using lectin-based ELISA assay and quantitative glycoproteomics analysis. *J. Proteome Res.* **12**, 3342–3352 (2013)
 30. Kuzmanov, U., Musrap, N., Kosanam, H., Smith, C.R., Batruch, I., Dimitromanolakis, A., Diamandis, E.P.: Glycoproteomic identification of potential glycoprotein biomarkers in ovarian cancer proximal fluids. *Clin. Chem. Lab. Med.* **51**, 1467–1476 (2012)
 31. Wang, P.H., Lee, W.L., Juang, C.M., Yang, Y.H., Lo, W.H., Lai, C.R., Hsieh, S.L., Yuan, C.C.: Altered mRNA expressions of sialyltransferases in ovarian cancers. *Gynecol. Oncol.* **99**, 631–639 (2005)
 32. Kuzmanov, U., Jiang, N., Smith, C.R., Soosaipillai, A., Diamandis, E.P.: Differential N-glycosylation of kallikrein 6 derived from ovarian cancer cells or the central nervous system. *Mol. Cell. Proteomics* **8**, 791–798 (2008)
 33. Berbeć, H., Paszkowska, A., Siwek, B., Gradziel, K., Cybulski, M.: Total serum sialic acid concentration as a supporting marker of malignancy in ovarian neoplasia. *Eur. J. Gynaecol. Oncol.* **20**, 389–392 (1999)
 34. Karlsson, N.G., McGuckin, M.A.: O-Linked glycome and proteome of high-molecular-mass proteins in human ovarian cancer ascites: identification of sulfation, disialic acid and O-linked fucose. *Glycobiology* **22**, 918–929 (2012)
 35. Santin, A.D., Ravindranath, M.H., Bellone, S., Muthugounder, S., Palmieri, M., O'Brien, T.J., Roman, J., Cannon, M.J., Pecorelli, S.: Increased levels of gangliosides in the plasma and ascitic fluid of patients with advanced ovarian cancer. *BJOG* **111**, 613–618 (2004)
 36. Goodarzi, M.T., Turner, G.A.: Decreased branching, increased fucosylation and changed sialylation of alpha-1 proteinase inhibitor in breast and ovarian cancer. *Clin. Chim. Acta* **236**, 161–171 (1995)
 37. Yue, K., Bian, M., Zhu, D., Liu, W., Siu, S.: Serum lipid-associated sialic acid (LSA) in diagnosing and monitoring ovarian cancer. *Zhongguo Yi Xue Ke Xue Yuan Xue Bao* **17**, 128–132 (1995)
 38. Petru, E., Sevin, B.U., Averette, H.E., Koehli, O.R., Perras, J.P., Hilsenbeck, S.: Comparison of three tumor markers—CA-125, lipid-associated sialic acid (LSA), and NB/70K—in monitoring ovarian cancer. *Gynecol. Oncol.* **38**, 181–186 (1990)
 39. Vardi, J.R., Tadros, G.H., Foemmel, R., Shebes, M.: Plasma lipid-associated sialic acid and serum CA 125 as indicators of disease status with advanced ovarian cancer. *Obstet. Gynecol.* **74**, 379–383 (1989)
 40. Strache, R.R., Büttner, H.H., Göcze, P.M., Briese, V.: Sialic acid concentrations in blood samples as well as in cyst and peritoneal fluid in patients with ovarian cysts and cystic ovarian tumors. *Zentralbl. Gynakol.* **112**, 1445–1453 (1990)
 41. Yang, L., Nyalwidhe, J.O., Guo, S., Drake, R.R., Semmes OJ: Targeted identification of metastasis-associated cell-surface sialoglycoproteins in prostate cancer. *Mol Cell Proteomics*. **10**(6): M110.007294 1–22, (2011).
 42. Saldo, R., Fan, Y., Fitzpatrick, J.M., Watson, R.W., Rudd, P.M.: Core fucosylation and alpha 2–3 sialylation in serum N-glycome is significantly increased in prostate cancer comparing to benign prostate hyperplasia. *Glycobiology* **21**, 195–205 (2011)
 43. PBassagañas, S., Pérez-Garay, M., Peracaula, R.: Cell surface sialic acid modulates extracellular matrix adhesion and migration in pancreatic adenocarcinoma cells. *Pancreas* **43**, 109–117 (2014)
 44. Mandal, C., Sarkar, S., Chatterjee, U., Schwartz-Albiez, R., Mandal, C.: Disialoganglioside GD3-synthase over expression inhibits survival and angiogenesis of pancreatic cancer cells through cell cycle arrest at S-phase and disruption of integrin-β1-mediated anchorage. *Int. J. Biochem. Cell Biol.* **53**, 162–173 (2014)
 45. Gruszewska, E., Chrostek, L., Cylwik, B., Tobolczyk, J., Szmikowski, M., Kuklinski, A., Kedra, B.: Serum sialic acid as a marker of pancreatic cancers. *Clin. Lab.* **59**, 781–788 (2013)
 46. Pérez-Garay, M., Arteta, B., Llop, E., Cobler, L., Pagès, L., Ortiz, R., Ferri, M.J., de Bolós, C., Figueras, J., de Llorens, R., Vidal-Vanaclocha, F., Peracaula, R.: α2,3-Sialyltransferase ST3Gal IV promotes migration and metastasis in pancreatic adenocarcinoma cells and tends to be highly expressed in pancreatic adenocarcinoma tissues. *Int. J. Biochem. Cell Biol.* **45**, 1748–1757 (2013)
 47. Yabu, M., Korekane, H., Takahashi, H., Ohigashi, H., Ishikawa, O., Miyamoto, Y.: Accumulation of free Neu5Ac-containing complex-type N-glycans in human pancreatic cancers. *Glycoconj. J.* **30**, 247–256 (2012)
 48. Masaki, Y., Oka, M., Ogura, Y., Ueno, T., Nishihara, K., Tangoku, A., Takahashi, M., Yamamoto, M., Irimura, T.: Sialylated MUC1 mucin expression in normal pancreas, benign pancreatic lesions, and pancreatic ductal adenocarcinoma. *Hepatogastroenterology* **46**, 2240–2245 (1999)
 49. Kameda, K., Shimada, H., Ishikawa, T., Takimoto, A., Momiyama, N., Hasegawa, S., Misuta, K., Nakano, A., Nagashima, Y., Ichikawa, Y.: Expression of highly polysialylated neural cell adhesion molecule in pancreatic cancer neural invasive lesion. *Cancer Lett.* **137**, 201–207 (1999)
 50. Liu, H., Ma, Q., Xu, Q., Lei, J., Li, X., Wang, Z., Wu, E.: Therapeutic potential of perineural invasion, hypoxia and desmoplasia in pancreatic cancer. *Curr. Pharm. Des.* **18**, 2395–2403 (2012)
 51. Amano, M., Eriksson, H., Manning, J.C., Detjen, K.M., André, S., Nishimura, S., Lehtiö, J., Gabius, H.J.: Tumour suppressor p16(INK4a) - anoikis-favouring decrease in N/O-glycan/cell surface sialylation by down-regulation of enzymes in sialic acid biosynthesis in tandem in a pancreatic carcinoma model. *FEBS J.* **279**, 4062–4080 (2012)
 52. Almaraz, R.T., Tian, Y., Bhattacharya, R., Tan, E., Chen, S.H., Dallas, M.R., Chen, L., Zhang, Z., Zhang, H., Konstantopoulos, K., Yarema, K.J.: Metabolic flux increases glycoprotein sialylation: implications for cell adhesion and cancer metastasis. *Mol. Cell. Proteomics* **11**, M112.017558 (2012)
 53. Maupin, K.A., Sinha, A., Eugster, E., Miller, J., Ross, J., Paulino, V., Keshamouni, V.G., Tran, N., Berens, M., Webb, C., Haab, B.B.: Glycogene expression alterations associated with pancreatic cancer epithelial-mesenchymal transition in complementary model systems. *PLoS ONE* **5**, e13002 (2012)
 54. Pérez-Garay, M., Arteta, B., Pagès, L., de Llorens, R., de Bolós, C., Vidal-Vanaclocha, F., Peracaula, R.: alpha2,3-sialyltransferase ST3Gal III modulates pancreatic cancer cell motility and adhesion in vitro and enhances its metastatic potential in vivo. *PLoS ONE*. **5**(9). (2010)
 55. Glavey, S.V., Maniery, S., Natoni, A., Sacco, A., Moschetta, M., Reagan, M.R., Murillo, L.S., Sahin, I., Wu, P., Mishima, Y., Zhang, Y., Zhang, W., Zhang, Y., Morgan, G., Joshi, L., Roccaro, A.M., Ghobrial, I.M., O'Dwyer, M.E.: The sialyltransferase ST3GAL6 influences homing and survival in multiple myeloma. *Blood* **124**(11), 1765–1776 (2014)
 56. Ilić, V., Milosević-Jovčić, N., Petrović, S., Marković, D., Stefanović, G., Ristić, T.: Glycosylation of IgG B cell receptor (IgG BCR) in

- multiple myeloma: relationship between sialylation and the signal activity of IgG BCR. *Glycoconj. J.* **25**, 383–392 (2008)
57. Fleming, S.C., Smith, S., Knowles, D., Skillen, A., Self, C.H.: Increased sialylation of oligosaccharides on IgG paraproteins—a potential new tumour marker in multiple myeloma. *J. Clin. Pathol.* **51**, 825–830 (1998)
 58. Zhao, Y., Li, Y., Ma, H., Dong, W., Zhou, H., Song, X., Zhang, J., Jia, L.: Modification of sialylation mediates the invasive properties and chemosensitivity of human hepatocellular carcinoma. *Mol. Cell. Proteomics* **13**, 520–536 (2014)
 59. Varki, A., Schauer, R.: Sialic Acids. In: Varki A, Cummings RD, Esko JD, et al., editors. *Essentials of glycobiology*. 2nd edition. Cold Spring Harbor (NY): Cold Spring Harbor Laboratory Press. Chapter 14. (2009)
 60. Pickup, J.C., Day, C., Bailey, C.J., Samuel, A., Chusney, G.D., Garland, H.O., Hamilton, K., Balment, R.J.: Plasma sialic acid in animal models of diabetes mellitus: evidence for modulation of sialic acid concentrations by insulin deficiency. *Life Sci.* **57**(1383–1391) (1995)
 61. Kopitzsch, S., Winkler, L., Oswald, B., Schlag, B., Dargel, R.: The sialylation rate of apolipoprotein E in insulin-dependent (IDDM) and non-insulin-dependent (NIDDM) diabetes mellitus. *Z. Med. Lab. Diagn.* **31**, 47–52 (1990)
 62. Nedić, O., Lagundžin, D., Masnikosa, R.: Posttranslational modifications of the insulin-like growth factor-binding protein 3 in patients with type 2 diabetes mellitus assessed by affinity chromatography. *J. Chromatogr. B Anal. Technol. Biomed. Life Sci.* **904**, 93–98 (2012)
 63. Waters, P.J., Flynn, M.D., Pennock, C.A., Corral, R.J., Greenwood, R.J., Eisenthal, R.: Decreased sialidase activity in mononuclear leucocytes of type 1 diabetic subjects: relationship to diabetic complications and glycaemic control. *Diabet. Med.* **12**, 670–673 (1995)
 64. Rogers, M.E., Williams, D.T., Nithyanathan, R., Rampling, M.W., Heslop, K.E., Johnston, D.G.: Decrease in erythrocyte glycophorin sialic acid content is associated with increased erythrocyte aggregation in human diabetes. *Clin. Sci. (Lond.)* **82**(309–313) (1992)
 65. Lee, C.L., Chiu, P.C., Pang, P.C., Chu, I.K., Lee, K.F., Koistinen, R., Koistinen, H., Seppälä, M., Morris, H.R., Tissot, B., Panico, M., Dell, A., Yeung, W.S.: Glycosylation failure extends to glycoproteins in gestational diabetes mellitus: evidence from reduced α 2-6sialylation and impaired immunomodulatory activities of pregnancy-related glycodelin-A. *Diabetes* **60**, 909–917 (2011)
 66. Kiljański, J., Ambroziak, M., Pachucki, J., Jazdzewski, K., Wiechno, W., Stachlewska, E., Górnicka, B., Bogdańska, M., Nauman, J., Bartoszewicz, Z.: Thyroid sialyltransferase mRNA level and activity are increased in Graves' disease. *Thyroid* **15**, 645–652 (2005)
 67. Janega, P., Cerná, A., Kholová, I., Brabencová, E., Babál, P.: Sialic acid expression in autoimmune thyroiditis. *Acta Histochem.* **104**, 343–347 (2002)
 68. Persani, L., Borgato, S., Romoli, R., Asteria, C., Pizzocaro, A., Beck-Peccoz, P.: Changes in the degree of sialylation of carbohydrate chains modify the biological properties of circulating thyrotropin isoforms in various physiological and pathological states. *J. Clin. Endocrinol. Metab.* **83**, 2486–2492 (1998)
 69. Babál, P., Janega, P., Cerná, A., Kholová, I., Brabencová, E.: Neoplastic transformation of the thyroid gland is accompanied by changes in cellular sialylation. *Acta Histochem.* **108**, 133–140 (2006)
 70. Krzeslak, A., Gaj, Z., Pomorski, L., Lipinska, A.: Sialylation of intracellular proteins of thyroid lesions. *Oncol. Rep.* **17**, 1237–1242 (2007)
 71. Takeyama, H., Kyoda, S., Okamoto, T., Manome, Y., Watanabe, M., Kinoshita, S., Uchida, K., Sakamoto, A., Morikawa, T.: The expression of sialic fibronectin correlates with lymph node metastasis of thyroid malignant neoplasms. *Anticancer Res.* **31**, 1395–1398 (2011)
 72. Vierbuchen, M., Schröder, S., Larena, A., Uhlenbruck, G., Fischer, R.: Native and sialic acid masked Lewis(a) antigen reactivity in medullary thyroid carcinoma. Distinct tumour-associated and prognostic relevant antigens. *Virchows Arch.* **424**, 205–211 (1994)
 73. Kökoğlu, E., Uslu, E., Uslu, I., Hatemi, H.H.: Serum and tissue total sialic acid as a marker for human thyroid cancer. *Cancer Lett.* **46**, 1–5 (1989)
 74. Jiang, M.S., Passaniti, A., Penno, M.B., Hart, G.W.: Adrenal carcinoma tumor progression and penultimate cell surface oligosaccharides. *Cancer Res.* **52**(8), 2222–2227 (1992)
 75. Dwivedi, C., Dixit, M., Hardy, R.E.: Plasma lipid-bound sialic acid alterations in neoplastic diseases. *Experientia* **46**, 91–94 (1990)
 76. Trouillas, J., Daniel, L., Guigard, M.P., Tong, S., Gouvernet, J., Jouanneau, E., Jan, M., Perrin, G., Fischer, G., Tabarin, A., Rougon, G., Figarella-Branger, D.: Polysialylated neural cell adhesion molecules expressed in human pituitary tumors and related to extrasellar invasion. *J. Neurosurg.* **98**, 1084–1093 (2003)
 77. Daniel, L., Trouillas, J., Renaud, W., Chevallier, P., Gouvernet, J., Rougon, G., Figarella-Branger, D.: Polysialylated-neural cell adhesion molecule expression in rat pituitary transplantable tumors (spontaneous mammatropic transplantable tumor in Wistar-Furth rats) is related to growth rate and malignancy. *Cancer Res.* **60**, 80–85 (2000)
 78. Ozyurt, E., Sönmez, H., Süer, S., Kökoğlu, E.: The prognostic importance of fibronectin and sialic acid levels in human pituitary adenomas. *Cancer Lett.* **100**, 151–154 (1996)
 79. Kökoğlu, E., Süer, S., Ozyurt, E., Siyahhan, A., Sönmez, H.: Plasma fibronectin and sialic acid levels in various types of human brain tumors. *Cancer Biochem. Biophys.* **15**, 35–40 (1995)
 80. Shen, Y., Kohla, G., Lrhorfi, A.L., Sipos, B., Kalthoff, H., Gerwig, G.J., Kamerling, J.P., Schauer, R., Tiralongo, J.: O-acetylation and de-O-acetylation of sialic acids in human colorectal carcinoma. *Eur. J. Biochem.* **271**, 281–290 (2004)
 81. Mann, B., Klussmann, E., Vandamme-Feldhaus, V., Iwersen, M., Hanski, M.L., Riecken, E.O., Buhr, H.J., Schauer, R., Kim, Y.S., Hanski, C.: Low O-acetylation of sialyl-Le(x) contributes to its over-expression in colon carcinoma metastases. *Int. J. Cancer* **72**, 258–64 (1997)